

Antidiarrheal Activity of *Albizia lebbeck* Leaves

Devendra S. Shirode^{1*}, Priyatama Powar¹, Ashwini Singh², Brijendra B. Jain³

ABSTRACT

Worldwide, more than 5–8 million deaths are caused each year by diarrhea. At present, used anti-diarrheal drugs show adverse effects in some form. Hence, there is a need to explore natural drugs as alternative therapy. Preliminary phytochemical screening, Acute oral toxicity and antidiarrheal activity of 70% ethanolic extract of the *Albizia lebbeck* leaves (ALL) were performed. ALL were assessed for antidiarrheal activity using castor oil, Prostaglandin-E₂ (PGE₂) and intestinal motility induced diarrhea in rats. On the basis of acute oral toxicity, 100 and 200 mg/kg body weight doses were selected for antidiarrheal activity. Preliminary phytochemical screenings revealed presence of flavonoids tannins and saponins in extract. ALL showed significant inhibitory activity against castor oil, PGE₂ induced diarrhea and intestinal motility. On the basis of the result, it can be concluded that the ALL possess significant antidiarrheal activity.

Keywords: *Albizia lebbeck*, Antidiarrheal activity, Castor oil, Charcoal, Ethanolic extract, Prostaglandin-E₂
Asian Pac. J. Health Sci., (2022); DOI: 10.21276/apjhs.2022.9.2.08

INTRODUCTION

Albizia lebbeck benth. (Mimosaceae) is a large, erect, unarmed and deciduous tree. On literature review it was found that, the leaves are used in ophthalmia.^[1] The bark is used in bronchial asthma and other allergic disorders.^[2] The flowers are useful in chronic cough and bronchitis.^[3] The seeds are aphrodisiac,^[1] useful in inflammation, scrofula, skin disease, leprosy, leukoderma, chronic catarrh, seminal weakness, ophthalmopathy, and poisoning.^[2] The leaves of the plant *A. lebbeck* are rich in flavon., echinocystic acid, β-sitosterol and vicenin II, etc.^[2] A native practitioner in this area had claimed that the leaves are highly useful in treating diarrhea. The modern literature revealed that the plant is reported to possess hepatoprotective,^[4] nootropic,^[5,6] anxiolytic,^[6] anticonvulsant,^[7,8] antifertility,^[9] antidiarrheal (seed),^[10] anti-inflammatory,^[11,12] and Antiulcer activity.^[13] There is no data reference available in the literature regarding the antidiarrheal activity of *A. lebbeck* leaves (ALL) either in humans or in any animal model. Hence, the present study was undertaken with the aim to assess the antidiarrheal activity of ALL so as to verify the claim of the native practitioner.

MATERIALS AND METHODS

Plant Material

The leaves of plant *A. lebbeck* were collected from fields of Anand, Gujarat. It was identified and authenticated by Prof. G.C. Jadeja, Dept of Agricultural Botany, Anand Agricultural University. The leaves were pulverized after shade dried at room temperature. The 70% ethanolic extract was prepared using 70% ethanol in a soxhlet apparatus after de-fatting with petroleum ether. Preliminary phytochemical investigation showed the presence of flavonoid and tannin in ALL. Hence, ALL was selected for the present activity.

Animals

Wistar albino rats (150–220 g) and mice (18–25 g) of either sex were used for the study. As per the Indian CPCSEA guidelines approved was obtained (1554/PO/a/11/CPCSEA).

¹Department of Pharmacology, Dr. D. Y. Patil College of Pharmacy, Pune, Maharashtra, India.

²Department of Patanjali Herbal Research, Patanjali Research Institute, Haridwar, Uttarakhand, India.

³Department of Pharmacology, Indrayani Institute of Pharmaceutical Education and Research, Pune, Maharashtra, India.

Corresponding Author: Dr. Devendra S. Shirode, Dr. D. Y. Patil College of Pharmacy, Pune, Maharashtra, India. Mobile: 7276790017. E-mail: devendrashirode@dyppharmaakurdi.ac.in

How to cite this article: Shirode DS, Powar P, Singh A, Jain BB. Antidiarrheal Activity of *Albizia lebbeck* Leaves. *Asian Pac. J. Health Sci.*, 2022;9(2):33-35.

Source of support: Nil

Conflicts of interest: None.

Received: 23/09/21 **Revised:** 12/10/21 **Accepted:** 10/11/21

Acute Toxicity Studies

As per fixed dose method of OECD Guide line no 420 given by CPCSEA, acute toxicity test was performed. Groups of 6 mice were administered test drug by oral route in the range of 2000–300 mg/kg and mortality was observed after 24 h.

Anti-diarrheal Activity

Castor oil-induced diarrhea^[14]

Animals were divided into four groups of six animals each and fasted for 18 h before the test with free access to water. The animals were treated as follows:

- Group I - Control (0.5 ml, 2% Gum acacia)
- Group II - Standard (loperamide 1 mg/kg i.p.)
- Group III - ALL (100 mg/kg p.o.)
- Group IV - ALL (200 mg/kg p.o.)

Each rat received 1 ml of castor-oil orally after 30 min treatment and housed separately in perforated cage over a clean filter paper. During 4 h, diarrheal episodes were observed, that is, onset of diarrhea, number of wet feces, number and mean weight of stool.

Gastro-intestinal motility test^[15]

The animals were divided into four groups of six rats each which were fasted for 24 h. The animals were treated as follows:

Group I - Control

Group II - Standard (Atropine sulfate 5 mg/kg i.m.)

Group III - ALL (100 mg/kg p.o.)

Group IV - ALL (200 mg/kg p.o.)

Each rat was received 1 ml of charcoal meal through oral route after 30 min of the above treatment. Thirty minutes after charcoal meal treatment, each rat was sacrificed and intestinal distance moved by the charcoal meal from pylorus to cecum was measured and expressed as a percentage of distance travelled from pylorus to cecum. The mean percentage movement of charcoal meal in ratio to the intestinal length, and percentage of inhibition was calculated by following the formula:

$$\% \text{Travelled} = \frac{\text{Distance travelled by the charcoal meal}}{\text{Total length of small intestine}} \times 100$$

$$\% \text{ of Inhibition} = \frac{\text{Total length of the} - \text{Distance travelled by small intestine the charcoal meal}}{\text{Total length of small intestine}} \times 100$$

Prostaglandin-E₂ (PGE₂) induced enteropooling model^[14]

The animals were divided into four groups of six animals each. These animals were placed in perforated cages and fasted for 18 h prior to the experiment and water ad libitum.

Treatment

Group I - Positive control (PGE₂ 100 µg/kg in 5% v/v ethanol in normal saline p.o.)

Group II - Standard (Loperamide 5 mg/kg i.p.)

Group III - ALL (100 mg/kg p.o.)

Group IV - ALL (200 mg/kg p.o.)

Group I was treated with PGE₂ and served as PGE₂ control. Immediately after the standard and extract treatment, each rat was administered PGE₂ in the Group II, III and IV. All the rats were killed after 30 min and the whole length of the intestine from the pylorus to the cecum was dissected out and its contents were weight and collected to measure its volume.

Statistical Analysis

The results were expressed as ±SEM. The statistical analysis was carried out by One-way Analysis of Variance followed by student "t" test.

RESULTS

In the castor oil induced diarrhea, the total number of feces, number of wet feces, and total weight of feces were reduced by test extract (ALL). Similarly, the time elapsed between the administration of the castor-oil and the excretion of the first diarrheic feces is increased, by 70% ALL. The test extract has shown dose dependent antidiarrheal activity against castor oil induced diarrhea in rats (Table 1). In the gastro intestinal motility model, the 70% ALL caused a decrease in propulsion of the charcoal meal through the gastrointestinal tract when compared to the control group. However, it was comparable to that of the standard drug (Table 2). In the PGE₂ induced enteropooling model, the test extract has shown

Table 1: Effect of 70% ethanolic extract of leaves of ALL on castor oil induced diarrhea in rats.

Gr. No.	Treatment	Onset of diarrhea (min)	Total number of feces	Number of wet feces	% of inhibition	Total wt. of feces (gm)	% of inhibition
I	Control	51.5±3.845	21.83±1.376	17.66±1.33	-	3.81±0.152	-
II	Loperamide 1 mg/kg	135.83±7.15***	5.83±0.60***	1.5±0.42***	91.50	0.77±0.42***	79.79
III	ALL 100 mg/kg	84.0±7.9 ^{ns}	11.33±1.05***	6.83±0.87**	61.32	1.85±0.14***	51.44
IV	ALL 200 mg/kg	103.83±11.39**	9.0±0.81***	3.83±0.60***	78.31	1.235±0.10***	67.58

Values are Mean±SEM., n=6. Significance ***P<0.001, **P<0.01, *P<0.05, ^{ns}P>0.05 (vs. Control) respectively. ALL: *Albizzia lebeck* leaves

Table 2: Effect of 70% ethanolic extract of leaves of ALL on gastrointestinal motility in rats

Gr. No.	Treatment	Mean Length of intestine (cms)	Mean distance travelled by charcoal (cms)	Mean percentage movement of charcoal (cms)	Percentage of inhibition (%)
I	Control	109.83±3.17	92.0±3.633	83.76	--
II	Atropine sulphate 5 mg/kg	110.36±3.56	47.28±3.48***	42.84	57.15
III	ALL 100 mg/kg. p.o.	113.33±3.48	78.26±2.07*	69.05	31.07
IV	ALL 200 mg/kg p.o.	116.83±4.88	70.86±1.40***	60.65	39.35

Values are Mean±SEM., n=6. Significance ***P<0.001, **P<0.01, *P<0.05, ^{ns}P>0.05 vs. Control. ALL: *Albizzia lebeck* leaves

Table 3: Effect of 70% ethanolic extract of leaves of ALL on PGE₂ induced enteropooling in rats

Gr. No.	Treatment	Dose	Mean volume of intestinal contents (ml)±SEM	% of inhibition	Mean weight of intestinal contents (ml) ± SEM	% of inhibition
I	Control	PGE ₂ 100µg/kg i.p.	3.05±0.260	--	3.77±0.22	--
II	Standard (loperamide)	5 mg/kg.p.o.	0.86±0.074***	71.86	1.69±0.15***	55.17
III	ALL	100 mg/kg. p.o.	2.11±0.113***	30.81	2.66±0.263***	29.44
IV	ALL	220 mg/kg.p.o.	1.48±0.197***	51.47	2.03±0.169***	46.56

ALL: *Albizzia lebeck* leaves, PGE₂: Prostaglandin-E₂. Values are Mean±SEM., n=6. Significance ***P<0.001, **P<0.01, *P<0.05, ^{ns}P>0.05 (vs. control), respectively

a significant inhibition of volume and weight of intestinal content in rats which was comparable to standard group animals (Table 3).

DISCUSSION

Castor oil contain ricinoleic acid cause permeability changes, irritation, and inflammation at intestinal mucosa^[16,17] lead to prostaglandins release.^[18,19] ALL delayed the onset of diarrhea in castor oil induced diarrhea. Anti-diarrheal activity of extracts may be due to inhibition of prostaglandins biosynthesis.

PGE₂ stimulate intestinal absorption of water and electrolytes.^[20] ALL showed inhibition of diarrhea may be due to inhibition of prostaglandins biosynthesis or by decreasing peristaltic movement.

In gastro-intestinal motility test, charcoal prevent absorption thereby decrease peristalsis movement.^[21] The inhibition of peristaltic movement with ALL may be due to antihistaminic and antianaphylactic activity.^[21,22]

However, further studies are needed to establish exact mechanism of action.

Phytochemical screening revealed the presence of tannins and flavonoids. Earlier studies showed that antidysentric and antidiarrheal properties of medicinal plants were due to tannins, alkaloids, saponins, flavonoids, sterol and/or triterpenes and reducing sugars.^[23] Hence, tannins and flavonoids may be responsible for the significant antidiarrheal activity in the present study. Further studies are required to identify and isolate the active principles responsible for the antidiarrheal activity to establish the exact mechanism of action of the test extract (ALL).

CONCLUSION

In conclusion, the present study demonstrates that ALL possesses antidiarrheal properties. In addition, the antidiarrheal activity may be attributed to the polyphenolic compounds of plant, namely tannins and flavonoids.

ACKNOWLEDGMENTS

We are thankful to Dr. D. Y. Patil College of Pharmacy, Akurdi, Pune for their constant support and providing all the facilities to carry out this research work.

REFERENCES

1. Kritikar KR, Basu BD. Indian Medicinal Plants. Vol. 2. Allahabad: Lalit Mohan Basu; 1998. p. 936-8.
2. Asolkar LV, Kakkar KK. Second Supplement to Glossary of Indian Medicinal Plants with Active Principles. New Delhi: Council of Scientific & Industrial Research; 1965. p. 36-8.
3. Vaidyaratnum PS. Variers: Indian Medicinal Plants. Vol. 2. Kottakkal: Arya Vaidyasala, Orient Longman; 1995. p. 81-4.
4. Shirode DS, Roy SP, Patel T, Rajendra SV, Jyothi TM, Setty SR. Evaluation of hepatoprotective effect of leaves of seventy percent ethanolic extract of *Albizia lebbek* in paracetamol induced experimental hepatic damage. *Plant Arch* 2008;8:797-801.
5. Chintawar SD, Somani RS, Kasture VS, Kasture SB. Nootropic activity of *Albizia lebbek* in mice. *J Ethnopharmacol* 2002;81:299-305.
6. Une HD, Sarveiya VP, Pal SC, Kasture VS, Kasture SB. Nootropic and anxiolytic activity of saponins of *Albizia lebbek* leaves. *Pharmacol Biochem Behav* 2001;69:439-44.
7. Kasture VS, Kasture SB, Pal SC. Anticonvulsant activity of *Albizia lebbek* leaves. *Indian J Exp Biol* 1996;34:78-80.
8. Kasture VS, Chopade CT, Deshmukh VK. Anticonvulsant activity of *Albizia lebbek*, *Hibiscus rosa-sinensis* and *Butea monosperma* in experimental animals. *J Ethnopharmacol* 2000;71:65-75.
9. Gupta RS, Kachhawa JB, Chaudhary R. Antifertility effects of methanol pod extract of *Albizia lebbek* (L) Benth in male rats. *Asian J Androl* 2004;6:155-9.
10. Besra SE, Gomes A, Chaudhury L, Vedasiromoni JR, Ganguly DK. Antidiarrhoeal activity of seed extract of *Albizia lebbek* Benth. *Phytother Res* 2002;16:529-33.
11. Pramanik KC, Bhattacharya P, Chatterjee TK, Mandal SC. Anti-inflammatory activity of methanol extract of *Albizia lebbek* bark. *Eur Bull Drug Res* 2005;13:71-5.
12. Shirode DS, Roy SP, Patel T, Rajendra SV, Jyothi TM, Setty SR. Antiinflammatory activity of ethanolic extract of *Albizia lebbek* leaves and *Madhuca longifolia* bark. *Int J Pharmacol Biol Sci* 2008;2:127-30.
13. Shirode DS, Patel T, Roy SP, Rajendra SV, Jyothi TM, Setty SR. Antilucer properties of 70 % ethanolic extract of leaves of *Albizia lebbek*. *Pharmacogn Mag* 2008;4:228.
14. Eakins KE, Sanner JM. Prostaglandins antagonists. In: Karim SM, editor. *Prostaglandins Progress in Research*. New York: Wiley-Interscience; 1972. p. 263-4.
15. Dajani EZ, Roge EA, Bertermann RE. Effects of E prostaglandins, diphenoxylate and morphine on intestinal motility *in vivo*. *Eur J Pharmacol* 1975;34:105-13.
16. Gaginella TS, Stewart JJ, Olson WA, Bass P. Actions of ricinoleic acid and structurally related fatty acid on the gastrointestinal tract II. Effects on water and electrolyte absorption *in vitro*. *J Pharmacol Exp Ther* 1975;195:355-61.
17. Ammon HV, Thomas PJ, Phillips SJ. Effects of oleic and ricinoleic acids on net jejunal water and electrolyte movement. *Perfusion studies in man*. *Clin Invest* 1974;53:374-9.
18. Pierce NF, Carpenter CJ, Elliott HZ, Greenough WB 3rd. Effects of prostaglandins, theophylline, and cholera exotoxin upon transmucosal water and electrolyte movement in the canine jejunum. *Gastroenterology* 1971;60:22.
19. Awouters F, Nimegeers CJE, Lenaerts FM, Janssen PA. Delay of castor oil diarrhoea in rats: A new way to evaluate inhibitors of prostaglandin biosynthesis. *J Pharm Pharmacol* 1978;30:41-5.
20. Jaffe BM. Prostaglandins and serotonin: Nonpeptide diarrhoeogenic hormones. *World J Surg* 1979;3:565-78.
21. Levy G. Gastrointestinal clearance of drugs with activated charcoal. *New Eng J Med* 1982;307:676-8.
22. Tripathi RM, Das PK. Studies on anti-asthmatic and anti-anaphylactic activity of *Albizia lebbek*. *Indian J Pharmacol* 1977;9:189-94.
23. Venkat RN, Chandraprakesh K, Shanta kumar BM. Pharmacological investigation of *Cardiospermum halicacabum* Linn in different animal models of diarrhea Indian. *J Pharmacol* 2006;5:346-9.